

A Distinct Acne Microbiome: Fact or Fiction?

REFERENCES

- Alexeyev OA, Jahns AC (2012) Sampling and detection of skin *Propionibacterium acnes*: current status. *Anaerobe* 18:479–83
- Alexeyev OA, Lundskog B, Ganceviciene R *et al.* (2012) Pattern of tissue invasion by *Propionibacterium acnes* in acne vulgaris. *J Dermatol Sci* 67:63–6
- Breternitz M, Flach M, Prassler J *et al.* (2007) Acute barrier disruption by adhesive tapes is influenced by pressure, time and anatomical location: integrity and cohesion assessed by sequential tape stripping. A randomized, controlled study. *Br J Dermatol* 156:231–40
- Fitz-Gibbon S, Tomida S, Chiu BH *et al.* (2013) *Propionibacterium acnes* strain populations in the human skin microbiome associated with acne. *J Invest Dermatol* 133: 2152–60
- Fredericks DN, Relman DA (1996) Sequence-based identification of microbial pathogens: a reconsideration of Koch's postulates. *Clin Microbiol Rev* 9:18–33
- Grice EA, Segre JA (2011) The skin microbiome. *Nat Rev Microbiol* 9:244–53
- Hall-Stoodley L, Costerton JW, Stoodley P (2004) Bacterial biofilms: from the natural environment to infectious diseases. *Nat Rev Microbiol* 2:95–108
- Jahns AC, Lundskog B, Ganceviciene R *et al.* (2012) An increased incidence of *Propionibacterium acnes* biofilms in acne vulgaris: a case-control study. *Br J Dermatol* 167: 50–8
- Kilian M, Scholz CF, Lomholt HB (2012) Multilocus sequence typing and phylogenetic analysis of *Propionibacterium acnes*. *J Clin Microbiol* 50:1158–65
- Lomholt HB, Kilian M (2010) Population genetic analysis of *Propionibacterium acnes* identifies a subpopulation and epidemic clones associated with acne. *PLoS One* 5:e12277
- McDowell A, Barnard E, Nagy I *et al.* (2012) An expanded multilocus sequence typing scheme for *Propionibacterium acnes*: investigation of 'pathogenic', 'commensal' and antibiotic resistant strains. *PLoS One* 7:e41480
- Mohammed D, Yang Q, Guy RH *et al.* (2012) Comparison of gravimetric and spectroscopic approaches to quantify stratum corneum removed by tape-stripping. *Eur J Pharm Biopharm* 82:171–4
- Pinkus H (1951) Examination of the epidermis by the strip method of removing horny layers. I. Observations on thickness of the horny layer, and on mitotic activity after stripping. *J Invest Dermatol* 16:383–6

A Distinct Acne Microbiome: Fact or Fiction?

Journal of Investigative Dermatology (2013) **133**, 2294–2295; doi:10.1038/jid.2013.259; published online 11 July 2013

TO THE EDITOR

We are pleased to see that Fitz-Gibbon *et al.* (2013) set out to conduct a prospective comparison of the microbiome of acne-prone and healthy skin using elegant molecular methods. They report that the skin of acne patients is enriched by particular strains, more specifically 16S ribotypes, with a very low prevalence in healthy subjects. The authors tell us that their study 'demonstrates a previously unreported paradigm of commensal strain populations that could explain the pathogenesis of human diseases', a powerful claim that requires robust evidence to support it.

The role of *Propionibacterium acnes* in the disease after which it was named has never been conclusively proven. It is not hard to see why, given that it is a ubiquitous resident of human skin from adrenarche to old age (Leyden *et al.*, 1975) and persists long after the acne-prone years. It is the dominant bacterial resident of healthy pilosebaceous follicles (Puhvel *et al.*, 1975). Recently, it has become possible to differentiate between different strains of *P. acnes* using techniques such as pulsed-field gel electrophoresis and multilocus sequence typing (Oprica *et al.*, 2004; Lomholt and Kilian,

2010; Kilian *et al.*, 2012; McDowell *et al.*, 2012). Although no typing study has been specifically designed to look for differences between strains from acne-prone and healthy skin, some data have been interpreted to suggest that differences may exist (Lomholt and Kilian 2010; McDowell *et al.*, 2012). Fitz-Gibbon *et al.* (2013) sought to specifically address this important question.

Detailed appraisal raises doubts about the authors' methods and the conclusions drawn from them. The sample population is stated to comprise 101 subjects, 49 acne patients and 52 healthy controls, all from South California. The reader is referred to the dbGaP website (http://www.ncbi.nlm.nih.gov/projects/gap/cgi-bin/study.cgi?study_id=phs000263.v1.p1) for further details, which show that the study sample initially comprised 148 subjects: 72 cases and 76 controls. No attempt has been made to match the age ranges of the patients and controls, and thus the oldest patient was 38 years old and the oldest control subject was 80 years old; eight control subjects were aged 42 years and above. Seven control subjects had been treated for acne in the past; it is unclear whether these had been excluded from the cohort

in the published article. Antibiotic therapy is known to perturb the microflora, and hence knowledge of previous treatment is pertinent; in 18 patients, treatment histories, even for the last 12 months, were not available. Among the cases, all had facial acne, but 48 (67%) did not have acne on the nose. In acne, the nose is often spared, as is the case here. Sampling from this largely unaffected site is therefore not advisable or likely to yield the evidence required to answer the question posed. Scant details are given of how the samples were collected and processed, but enough to show that they were not ideal for comparing acne versus control subjects. Pore strips were used to obtain pooled follicular casts from the nose. In both acne and non-acne, predominantly normal follicles will have been sampled by this procedure, as they always vastly outnumber lesional ones. It is doubtful whether this method will adequately sample inflamed lesions even in the minority of patients in whom they may have been present.

The text refers to the pore tape samples as microcomedones and goes on to say that individual microcomedones were isolated using forceps. Acne is characterized by comedones of several types, of which whiteheads (closed comedones), blackheads (open comedones), and macrocomedones are the most common types (Cunliffe *et al.*, 2000). Microcomedones

Abbreviation: RT, ribotype

Accepted article preview online 12 June 2013; published online 11 July 2013

in situ are not visible to the naked eye and can only be seen in electron micrographs of skin biopsies. Pore stripping, like cyanoacrylate glue, removes vellus hairs to which follicular casts are attached. These casts comprise cellular debris, sebum, and bacteria, and are typically larger when derived from acne-prone follicles. Casts from acne patients may or may not include microcomedones depending on the site sampled; those from healthy skin are certainly not microcomedones. On the nose, trichostasis spinulosa (accumulation of shed vellus hairs in the follicular lumen, Harford *et al.*, 1996) is frequently present in acne and non-acne; affected follicles will be sampled by pore stripping. It is not apparent whether the skin surface was sterilized before use of the pore tape and, as the entire tape was processed (except in the metagenomics), transient surface and resident follicular bacteria may have been included.

The results of this study show that six ribotypes were more abundant in acne patients than in controls, four significantly so. Of these six ribotypes, three (RT4, RT5, and RT10) carry a G1058C nucleotide substitution associated with resistance to tetracyclines (Ross *et al.*, 1998; Oprica *et al.*, 2005). The presence of these tetracycline-resistant variants on acne-prone skin is almost certainly a reflection of selective pressure, and not virulence. Oral tetracyclines have been a popular acne therapy for over 50 years and are still widely prescribed. Chronic antibiotic administration may or may not have created them, but it put them at a competitive advantage over susceptible

lineages. Outside the United States of America, most isolates of *P. acnes* from acne patients remain tetracycline susceptible, especially in countries where tetracyclines for acne are less intensively used (Ross *et al.*, 2003; Oprica *et al.*, 2004; Luk *et al.*, 2013). In the United States of America, *P. acnes* resistant to anti-acne antibiotics were sought but not found on the skin of 1,000 acne patients sampled in the mid 1970s (Leyden, 1976); however, they were detected in the following decade (Leyden *et al.*, 1983).

The possibility that strains of *P. acnes* specifically associated with acne exist is an exciting one. Demonstrating that this is true would be a first step toward determining whether some strains are more pathogenic than others or are better able to survive within acne lesions. The authors' sophisticated molecular analyses are well suited to providing the answers as long as more appropriate samples are used.

CONFLICT OF INTEREST

The authors state no conflict of interest.

E. Anne Eady¹ and Alison M. Layton¹

¹Department of Dermatology, Harrogate and District NHS Foundation Trust, Harrogate, UK
E-mail: eaeady@gmail.com

REFERENCES

- Cunliffe WJ, Holland DB, Clark SM *et al.* (2000) Comedogenesis: some new aetiological, clinical and therapeutic strategies. *Br J Dermatol* 142:1084–91
- Fitz-Gibbon S, Tomida S, Chiu BH *et al.* (2013) *Propionibacterium acnes* strain populations in the human skin microbiome associated with acne. *J Invest Dermatol* 133:2147–55
- Harford RR, Cobb MW, Miller ML (1996) Trichostasis spinulosa: a clinical simulant of acne open comedones. *Pediatr Dermatol* 13:490–2
- Kilian M, Scholz CF, Lomholt HB (2012) Multilocus sequence typing and phylogenetic analysis of *Propionibacterium acnes*. *J Clin Microbiol* 50:1158–65
- Leyden JJ (1976) Antibiotic resistant acne. *Cutis* 17:593–6
- Leyden JJ, McGinley KJ, Mills OH *et al.* (1975) Age-related changes in the resident bacterial flora of the human face. *J Invest Dermatol* 65:379–81
- Leyden JJ, McGinley KJ, Cavalieri S *et al.* (1983) *Propionibacterium acnes* resistance to antibiotics in acne patients. *J Am Acad Dermatol* 8:41–5
- Lomholt HB, Kilian M (2010) Population genetic analysis of *Propionibacterium acnes* identifies a subpopulation and epidemic clones associated with acne. *PLoS ONE* 5:e12277
- Luk NM, Hui M, Lee HC *et al.* (2013) Antibiotic-resistant *Propionibacterium acnes* among acne patients in a regional skin centre in Hong Kong. *J Eur Acad Dermatol Venereol* 27:31–6
- McDowell A, Barnard E, Nagy I *et al.* (2012) An expanded multilocus sequence typing scheme for *Propionibacterium acnes*: investigation of 'pathogenic', 'commensal' and antibiotic resistant strains. *PLoS One* 7:e41480
- Oprica C, Emtestam L, Lapins J *et al.* (2004) Antibiotic-resistant *Propionibacterium acnes* on the skin of patients with moderate to severe acne in Stockholm. *Anaerobe* 10:155–64
- Oprica C, Lofmark S, Lund B *et al.* (2005) Genetic basis of resistance in *Propionibacterium acnes* strains isolated from diverse types of infection in different European countries. *Anaerobe* 11:137–43
- Puhvel SM, Reisner RM, Amirian DA (1975) Quantification of bacteria in isolated pilosebaceous follicles in normal skin. *J Invest Dermatol* 65:525–31
- Ross JI, Eady EA, Cove JH *et al.* (1998) 16S rRNA mutation associated with tetracycline resistance in a gram-positive bacterium. *Antimicrob Agents Chemother* 42:1702–5
- Ross JI, Snelling AM, Carnegie E *et al.* (2003) Antibiotic-resistant acne: lessons from Europe. *Br J Dermatol* 148:467–78

Response to the Commentaries on the Paper: *Propionibacterium acnes* Strain Populations in the Human Skin Microbiome Associated with Acne

Journal of Investigative Dermatology (2013) 133, 2295–2297; doi:10.1038/jid.2013.275; published online 11 July 2013

TO THE EDITOR

In our recent paper, we reported our metagenomic study of the skin

microbiome associated with acne (Fitz-Gibbon *et al.*, 2013). Using 16S ribosomal DNA (rDNA) analysis and

whole-genome sequence analysis of 71 *Propionibacterium acnes* strains, we demonstrated that the strain populations of *P. acnes* in pilosebaceous units were distinct between acne